



# Increased pulmonary serotonin transporter in patients with chronic obstructive pulmonary disease who developed pulmonary hypertension

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## Abstract

**Purpose** Pulmonary hypertension (PH) is characterized by a progressive remodelling of the pulmonary vasculature resulting in right heart failure and eventually death. The serotonin transporter (SERT) may be involved in the pathogenesis of PH in patients with chronic-obstructive pulmonary disease (COPD). This study investigated for the first time the SERT in vivo availability in the lungs of patients with COPD and PH (COPD+PH).

**Methods** SERT availability was assessed using SERT-selective [<sup>11</sup>C]DASB and positron emission tomography/computed tomography (PET/CT) with dynamic acquisition over 30 min in 4 groups of 5 participants each: COPD, COPD+PH, pulmonary arterial hypertension, and a healthy control (HC). Time activity curves were generated based on a volume of interest within the middle lobe. Tissue-to-blood concentration ratios after 25 to 30 min (TTBR<sub>25–30</sub>) served as receptor parameter for group comparison and were corrected for lung tissue attenuation. Participants underwent comprehensive pulmonary workup. Statistical analysis included group comparisons and correlation analysis.

**Results** [<sup>11</sup>C]DASB uptake peak values did not differ among the cohorts after adjusting for lung tissue attenuation, suggesting equal radiotracer delivery. Both the COPD and COPD+PH cohort showed significantly lower TTBR<sub>25–30</sub> values after correction for lung attenuation than HC. Attenuation corrected TTBR<sub>25–30</sub> values were significantly higher in the COPD+PH cohort than those in the COPD cohort and higher in non-smokers than in smokers. They positively correlated with invasively measured severity of PH and inversely with airflow limitation and emphysema. Considering all COPD patients ± PH, they positively correlated with right heart strain (NT-proBNP).

**Conclusion** By applying [<sup>11</sup>C]DASB and PET/CT, semiquantitative measures of SERT availability are demonstrated in the lung vasculature of patients with COPD and/or PH. COPD patients who developed PH show increased pulmonary [<sup>11</sup>C]DASB uptake compared to COPD patients without PH indicating an implication of pulmonary SERT in the development of PH in COPD patients.

**Keywords** Serotonin transporter · Positron emission tomography · Computed tomography · Chronic obstructive pulmonary disease · Pulmonary hypertension

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Armin Frille, Michael Rullmann, Swen Hesse and Hans-Juergen Seyfarth contributed equally to this work.

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## Introduction

Pulmonary hypertension (PH) is a hemodynamic disorder that affects both the respiratory and cardiovascular system and may cause multiple clinical conditions [1]. According to the current guidelines, PH is defined as an increase in mean pulmonary arterial pressure (PAPm)  $\geq$  25 mmHg at rest as assessed by right heart catheterization (RHC) [1]. It is characterized by a vasculopathy of the small pulmonary arteries that comprises vasoconstriction and proliferation in all layers of the vessel wall as well as fibrosis and inflammation. It may be complicated by right failure and eventually death. The clinical classification of PH categorizes multiple clinical conditions into five groups: Briefly, PH can be due to (1) pulmonary arterial hypertension (PAH) including idiopathic, familial, drug, and toxin-induced and associated forms, (2) left heart diseases, (3) lung diseases and/or hypoxia, for example, chronic obstructive lung diseases (COPD) and interstitial lung diseases, (4) chronic thromboembolic PH (CTEPH), and (5) unclear multifactorial mechanisms (hematologic, systemic, or metabolic disorders).

Thereof, left heart diseases and lung diseases and/or hypoxia are the most prevalent clinical conditions in PH. The presence of PH in COPD patients is a strong predictor of mortality in COPD [2]. Pathophysiologically, chronic inflammation response due to chronic inhalation of cigarette smoke or other noxious particles induces airway limitation and irreversible parenchymal lung tissue destruction resulting in emphysema, as a hallmark of advanced stage of COPD [3]. As a consequence thereof, pulmonary vascular bed becomes rarefied affecting pulmonary perfusion. The resulting alveolar hypoxia additionally causes hypoxic pulmonary vasoconstriction (HPV), which altogether leads to the development of PH in COPD (COPD+PH).

A role of serotonin, or 5-hydroxytryptamine (5-HT), in the development of PH has been proposed for many decades due to serotonin's vasoconstrictive and proliferative properties leading to the so-called serotonin hypothesis of PH [4]. Evidence suggests that pulmonary endothelial cells from PAH patients overexpress tryptophan hydroxylase 1 (TPH1) leading to increased endothelial serotonin synthesis and secretion towards pulmonary arterial smooth muscle cells (PASMC) [5]. Serotonin induces relevant vasoconstriction in the pulmonary arteries via its receptors (e.g., 5-HT<sub>1B</sub>) and proliferation of PASMCs via both its receptors and transporters (Fig. 1), subsequently contributing to the development or aggravation of PH [6–8]. Increased serotonin signalling has been implicated in the development of PAH after the use of antidepressants in pregnant women giving birth to new-borns

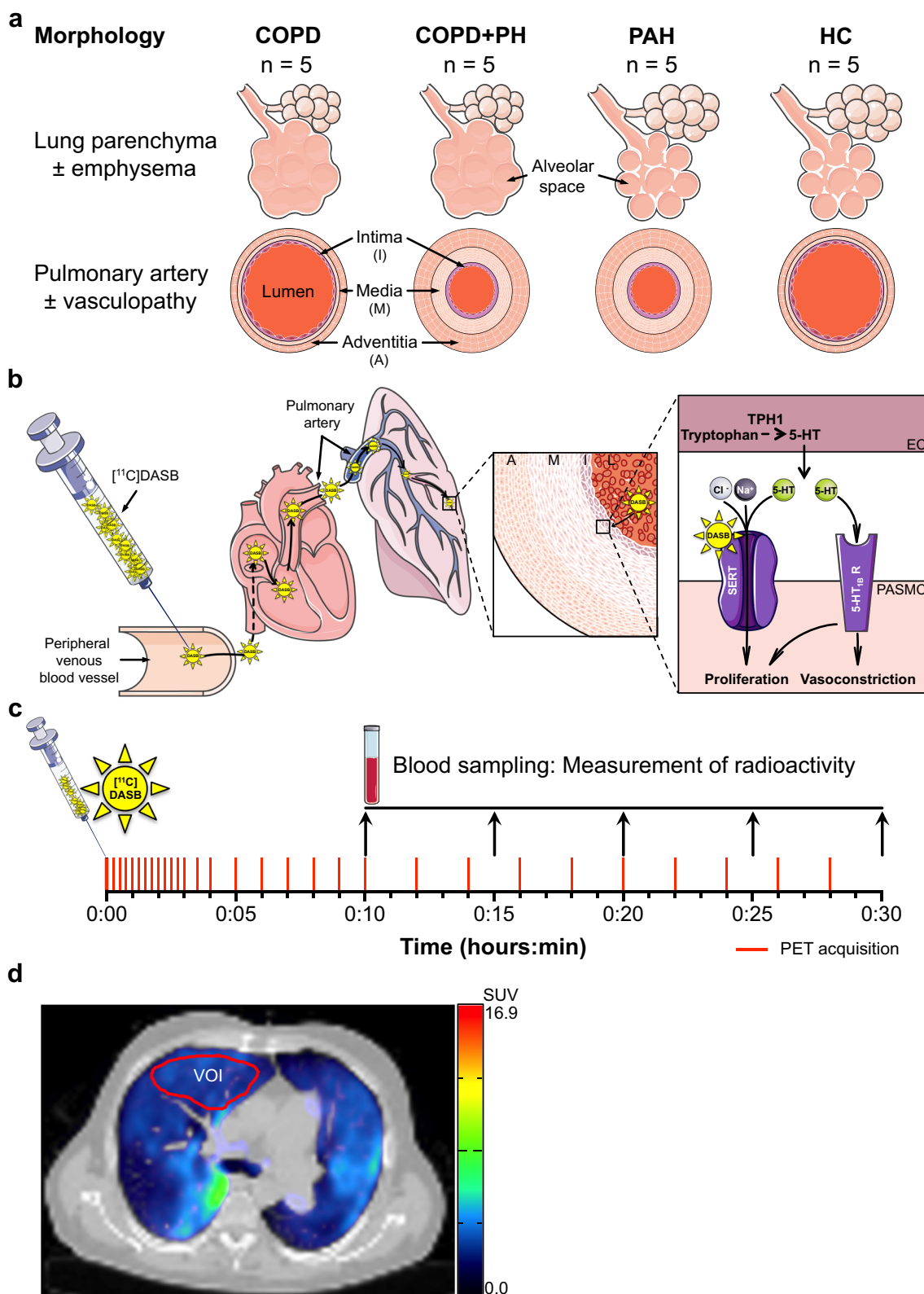
**Fig. 1** Artwork showing study design and study procedures. (a) Description of cohort's characteristics ( $\pm$  emphysema,  $\pm$  vasculopathy). (b) Pathway of [<sup>11</sup>C]DASB through the vascular system with binding to serotonin transporter (SERT) on the pulmonary artery smooth muscle cell (PASMC) surface. (c) Time points of PET acquisitions and blood sampling after [<sup>11</sup>C]DASB administration (d) Representative coregistered transversal PET/CT image of a patient with COPD and PH showing a manually and click-wise selected volume of interest (VOI) in the middle lobe of the lung. On right side of the PET/CT image, a color scale shows the range of SUV values. 5-HT 5-hydroxytryptamine, A adventitia, CT chloride ion, COPD chronic obstructive pulmonary disease, EC endothelial cell, HC healthy control, I intima, L lumen, n number of participants, Na<sup>+</sup> sodium ion, M media, min minute, PAH pulmonary arterial hypertension, PASMC pulmonary arterial smooth muscle cells, PH pulmonary hypertension, SUV standardized uptake value, TPH1 tryptophan hydroxylase 1

with persistent PH and appetite suppressants as they act as selective serotonin reuptake inhibitors (SSRI) or serotonin receptor agonists [1]. Moreover, in PAH patients, incident SSRI use was associated with increased mortality and a greater risk of clinical worsening [9].

The serotonin transporter (SERT) is a Na<sup>+</sup>/Cl<sup>−</sup>-coupled symporter for the biogenic amine serotonin and functions to reduce extracellular levels of serotonin, as it does on the presynaptic nerve terminals in the central nervous system [10]. The lung has been shown to play a major role in the physiologic removal of circulating serotonin through pulmonary SERT [11–13]. Increased expression or activity of SERT in the PASMC has been observed in patients with PAH and COPD+PH [6, 14]. Consistent with this observation is the fact that hypoxic SERT knockout mice develop less severe PH and vascular remodelling than the wild-type counterpart [15] emphasizing the role of SERT in the pathophysiology of PH under hypoxic conditions, such as in COPD patients.

Positron emission tomography (PET) studies using an <sup>11</sup>C-labelled antidepressant directed to target SERT found high accumulation in the lung [16]. This suggests that the lung functions as a reservoir for antidepressants via binding to SERT. The SERT-selective radioligand [<sup>11</sup>C]DASB, which is a sulfanyl-benzonitrile derivate, is clinically used by means of PET and computed tomography (PET/CT) imaging in the fields of schizophrenia, epilepsy, and depression [17]. Moreover, [<sup>11</sup>C]DASB and PET imaging also demonstrated pulmonary SERT availability in humans [18], and the intake of SERT-selective paroxetine reduced the lung uptake of a radioligand selective for monoamine transporter and SERT ([<sup>123</sup>I]FP-CIT) as compared to placebo intake as shown by scintigraphy [19].

In here, we carried out a first-in-human pilot study using semiquantitative uptake measures of the SERT-selective radioligand [<sup>11</sup>C]DASB to assess whether the



serotonin pathway in the lung is pathophysiologically relevant for the development of PH in patients with COPD. To this end, we hypothesized that COPD patients

who developed PH (COPD+PH) show increased pulmonary SERT availability compared with COPD patients without PH.

## Methods

### Study design and subjects

We performed a prospective pilot study of 15 patients with COPD, COPD+PH, or PAH, and five healthy controls (HC) (Fig. 1(a)). We selected these participants from the inpatient and outpatient clinic of our Department of Respiratory Medicine. Inclusion criteria consisted of parameters from the respiratory workup defining advanced stage of COPD and/or PH, respectively. Participants not meeting the criteria for advanced disease stage were not included. As a control group, we recruited healthy volunteers, who were matched to age and were non-smokers.

### Clinical, laboratory, and hemodynamic assessment

The diagnoses for COPD, COPD+PH, and PH were established according the current clinical guidelines [1, 20]. The respiratory workup for participants comprised pulmonary function test (PFT) including spirometry and body plethysmography, 6-min walking distance (6MWD) including the Borg score for rating dyspnoea (0 points: no dyspnoea; 10 points: maximal dyspnoea), capillary blood gas analysis (BGA), diffusing capacity of the lung for carbon monoxide (DLCO), serum levels of N-terminal pro-brain natriuretic peptide (NT-proBNP), as well as the calculation of the oxygenation ratio (arterial oxygen tension [ $\text{PaO}_2$ ]/inspiratory oxygen fraction [ $\text{FiO}_2$ ]), and alveolar-arterial gradient ( $\text{A-aO}_2$ ). Disease severity and risk of death estimates for COPD patients were assessed using the multidimensional BODE score that includes body mass index (B), airflow obstruction (O), dyspnoea (D), and exercise capacity (E) [21]. Right heart catheterization (RHC) provided invasively measured parameters on mean pulmonary arterial pressure (PAPm), pulmonary vascular resistance (PVR), and cardiac index (CI), establishing the diagnosis of PH and determining its severity. Severity of PH was clinically evaluated using the World Health Organization functional class (WHO-FC) [22]. Care has been taken that none of the participants were under SSRI or monoamine oxidase (MAO) inhibitors therapy during study enrolment.

### [ $^{11}\text{C}$ ]DASB PET/CT

Pulmonary SERT availability was assessed after intravenous injection of SERT-selective [ $^{11}\text{C}$ ] 3-amino-4-(2-dimethylaminomethylphenylsulfanyl)-benzonitrile ([ $^{11}\text{C}$ ]DASB) with averaged  $495 \pm 6$  MBq using an integrated PET/CT scanner (Biograph 16 PET/CT Scanner [Siemens Medical Solutions, Erlangen, Germany]). The dynamic acquisition was performed over 30 minutes (min) and comprised 30 measuring points ( $12 \times 15$  s [seconds],  $2 \times 30$  s,  $6 \times 60$  s,  $10 \times 120$  s). A low-dose CT of the lung was performed in each

subject for anatomic coregistration and attenuation correction. The radioactivity in blood was measured at 10, 15, 20, 25, and 30-min post injection (p.i.) in each participant using a Cobra gamma counter (Packard Instrument Company, Meriden, CT, USA) and decay was corrected to the start time of the PET/CT scan (Fig. 1(b, c)).

### Image analysis

Time activity curves (TACs) were generated from each of the 30 measuring points based on a volume of interest (VOI), which were manually defined within the middle lobe of the lung, averaging  $14.0 \pm 4.7$  mm<sup>3</sup> without intergroup differences (Fig. 1(d)). Standardized uptake values (SUV) were computed from tracer activity measured with PET according to  $\text{SUV} = \text{tracer-activity-in-tissue}/(\text{injected-dose/body-weight})$ . Both the maximum standardized uptake value ( $\text{SUV}_{\text{max}}$ ) after 0 to 3 min and the SUV after 25 to 30 min ( $\text{SUV}_{25-30}$ ) served as model-free parameter for group comparison. Tissue-to-blood (TTBR) and tissue-to-plasma (TTPR) concentration ratios were calculated by means of the averaged activity of the last two blood or plasma samples (25–30 min p.i.), respectively.

### Statistical analysis

Data were compiled and their distributions were estimated (histogram analysis, skewness, kurtosis, Shapiro-Wilk test). Group differences were calculated using the Student's *t* test or Mann-Whitney *U* test for comparison of 2 groups and one-way analysis of variance (ANOVA) followed by Tukey's post hoc correction for comparison of > 2 groups. Analysis of covariance (ANCOVA) was performed to adjust the TTBR and TTPR values for the mean lung tissue attenuation of the middle lobe. The calculation of partial Spearman rank correlation coefficient between  $\text{TTBR}_{25-30}$  (independent variable) and clinical or hemodynamic parameters (dependent variables) was used to adjust for lung tissue attenuation (control variable). Statistical significance was accepted at a level of a two-sided  $P < 0.05$ . Results are expressed as mean  $\pm$  standard deviation (SD) or 95% confidence interval (CI). Data analysis, calculation, and preparation of figures were conducted using the software package GraphPad Prism (v8.3.0 for macOS, La Jolla, California, USA), SPSS (v25.0, IBM Corporation, Chicago, IL, USA), Matlab (v7.13, The MathWorks Inc., Natick, MA, USA), and R: A Language and Environment for Statistical Computing (v3.4, R Foundation for Statistical Computing, Vienna, Austria, 2017, <http://www.R-project.org>). Figure 1 contains modified graphic content provided by Servier Medical Art by Servier (<https://smart.servier.com>) licensed under a Creative Commons Attribution 3.0 unported license (CC BY 3.0).



## Results

### Characteristics of subjects

A coregistered [ $^{11}\text{C}$ ]DASB using PET/CT with dynamic acquisition over 30 min and a respiratory workup was performed in 19 participants (1 participant was not measured by PET/CT for technical reasons). In addition, participants underwent hemodynamic assessment by means of RHC, except for the HC. All PAH patients received a drug combination therapy consisting of an endothelin receptor antagonist (ERA) plus a phosphodiesterase type 5 inhibitor (PDE-5i) (3/5) or a guanylate cyclase stimulator (2/5) and an inhaled prostacyclin analogue (1/5). In COPD+PH cohort, 3/5 participants received a specific drug therapy, of which 2 were on monotherapy (PDE-5i) and 1 on combination therapy (PDE-5i plus ERA).

Table 1 gives an overview on the clinical, hemodynamic, laboratory, and radiologic characteristics of the participants. Age and body mass index (BMI) were equally distributed among the 4 cohorts. All COPD patients with or without PH exhibited severe airflow limitation while COPD patients without PH significantly expressed a higher GOLD stage ( $P < 0.05$ ) and higher BODE score ( $P < 0.05$ ) than COPD+PH patients (Supplementary Fig. 1). COPD patients showed stronger airflow limitation (lower FEV<sub>1</sub> values, higher total airway resistance [ $R_{\text{tot}}$ ] values), higher ratio values of residual volume/total lung capacity (RV/TLC) as a surrogate for emphysema and poorer diffusing capacity (lower DLCO values) as compared to the PAH and HC cohorts. Additional analysis of clinical and hemodynamic characteristics of COPD patients with or without PH is shown in the Supplementary Fig. 2.

### Pulmonary SERT was quantifiable using [ $^{11}\text{C}$ ]DASB PET/CT

Pulmonary uptake of SERT-selective [ $^{11}\text{C}$ ]DASB was quantified using PET/CT in all cohorts investigated (Fig. 2). TACs of all cohorts show initial peak of SUV, designating the pulmonary perfusion and radiotracer delivery and a subsequent decline in SUV for the rest of the acquisition time.

### Pulmonary perfusion in COPD did not affect pulmonary [ $^{11}\text{C}$ ]DASB uptake

The lung tissue attenuation measured by means of CT among the cohorts only differed in the ANOVA, but not in the post hoc comparisons (Fig. 3(a)). Both TTBR<sub>max</sub> and TTPR<sub>max</sub> corrected for lung attenuation (ANCOVA) did not differ between the cohorts (Fig. 3(b)).

### Pulmonary [ $^{11}\text{C}$ ]DASB uptake significantly differed between COPD patients with and without PH

Both the COPD and COPD+PH cohort showed significantly lower attenuation corrected TTBR<sub>25–30</sub> and TTPR<sub>25–30</sub> values than the HC (Fig. 3(c)). Attenuation corrected TTBR<sub>25–30</sub> values were significantly higher in COPD+PH than in COPD patients without PH ( $P = 0.038$ ), while using attenuation corrected TTPR<sub>25–30</sub> values, this group comparison confirmed the trend but did not reach, if only just, the significance level ( $P = 0.054$ ). Both attenuation corrected TTBR<sub>25–30</sub> and TTPR<sub>25–30</sub> did not differ between the HC and PAH cohort ( $P = 0.243$ ,  $P = 0.172$ , respectively). Smokers showing long-term exposure (averaging 43 pack years) were measured significantly lower TTBR<sub>25–30</sub> values than the non-smokers ( $5.7 \pm 2.8$  vs.  $12.7 \pm 5.2$ ,  $P = 0.005$ ) (Fig. 3(d)).

### Pulmonary [ $^{11}\text{C}$ ]DASB uptake significantly correlated with clinical and hemodynamic parameters

Partial Spearman rank correlation analysis of TTBR<sub>25–30</sub> was performed with clinical and hemodynamic parameters to adjust for lung tissue attenuation (Fig. 4, Table 2). With regards to severity of PH, attenuation corrected TTBR<sub>25–30</sub> values positively correlated with PAPm and PVR ( $\rho = 0.69$  and  $\rho = 0.65$ , respectively). In terms of severity of COPD, TTBR<sub>25–30</sub> values that were corrected for lung attenuation positively correlated with FEV<sub>1</sub>/FVC ( $\rho = 0.72$ ), as an indicator of airflow limitation, and inversely with RV/TLC ( $\rho = -0.80$ ), as an indicator of the degree of pulmonary emphysema. Exercise capacity evaluated by 6MWD positively correlated with TTBR<sub>25–30</sub> values corrected for lung attenuation ( $\rho = 0.79$ ). Concerning the pulmonary vascular mismatch, attenuation corrected TTBR<sub>25–30</sub> positively correlated with the oxygenation ratio, i.e., PaO<sub>2</sub>/FiO<sub>2</sub> ( $\rho = 0.74$ ) and with the diffusing capacity for carbon monoxide, i.e., DLCO ( $\rho = 0.77$ ) and negatively with hypercapnia, i.e., PaCO<sub>2</sub>, measured by BGA ( $\rho = -0.68$ ).

## Discussion

To the best of our knowledge, this is the first prospective trial to study the pulmonary SERT availability in COPD patients having developed PH by using [ $^{11}\text{C}$ ]DASB and PET/CT. By applying a semiquantitative approach, we found that [ $^{11}\text{C}$ ]DASB was not only measurable in the lung parenchyma of the 4 cohorts tested; its uptake also differed significantly between participants with COPD and with COPD+PH. Pulmonary SERT availability correlated positively with the severity of PH, arterial oxygenation (PaO<sub>2</sub>/FiO<sub>2</sub>), and exercise capacity (6MWD) as well as inversely with airflow limitation, emphysema, and carbon dioxide retention. Furthermore,

**Table 1** Characteristics of participants

Characteristic	COPD	COPD+PH	PAH	HC
Participants, <i>n</i>	5	5	5	5
Sex, male/female	4/1	4/1	0/5	2/3
Age (years)	57.2 ± 2.8	55.8 ± 9.9	57.0 ± 2.8	56.2 ± 8.0
BMI (kg × m <sup>-2</sup> )	25.5 ± 3.2	21.6 ± 2.3	24.6 ± 4.7	26.3 ± 4.1
Smokers, <i>n</i> , pys	5, 50 (30–60)	3, 30 (20–50)	0	0
BODE score <sup>‡</sup>	7 (7–9)	5 (3–7)	-	-
WHO-FC	-	3 (3–4)	3 (3–4)	-
Pulmonary function test				
FEV <sub>1</sub> (% predicted) <sup>‡</sup>	22.1 ± 4.1	46.3 ± 17.3	79.6 ± 16.0	98.7 ± 12.8
COPD stage	4	3 (2–4)	-	-
<i>R</i> <sub>tot</sub> (% predicted) <sup>‡</sup>	508.3 ± 285.3	168.2 ± 87.0	81.2 ± 26.9	60.2 ± 19.9
RV/TLC (%) <sup>‡</sup>	57.7 ± 2.7	62.5 ± 10.8	36.2 ± 8.1	44.4 ± 11.3
DLCO (% predicted) <sup>‡</sup>	16.9 ± 13.5	18.9 ± 6.3	55.2 ± 19.8	-
Blood gas analysis*				-
PaCO <sub>2</sub> (mm Hg) <sup>‡</sup>	44.3 ± 2.1	33.1 ± 5.5	29.4 ± 1.7	-
PaO <sub>2</sub> (mm Hg)	56.3 ± 4.8	52.8 ± 9.7	68.6 ± 16.5	-
PaO <sub>2</sub> /FiO <sub>2</sub> (mm Hg)	231.1 ± 46.5	264.0 ± 51.0	323.8 ± 75.8	-
SaO <sub>2</sub> (%)	89.4 ± 2.1	91.2 ± 3.5	88.3 ± 11.2	-
A-aO <sub>2</sub> (mm Hg)	35.9 ± 3.3	49.5 ± 11.4	43.9 ± 21.2	-
Exercise capacity				-
6MWD (m) <sup>‡</sup>	205.6 ± 101.3	341.0 ± 72.5	424.0 ± 128.4	-
Need for O <sub>2</sub> at rest, <i>n</i> (%)	5 (100)	4 (80)	1 (20)	-
O <sub>2</sub> flow (L × min <sup>-1</sup> ) <sup>†</sup>	4 (2–10)	4 (0–5)	0 (0–4)	-
Borg score <sup>‡</sup>	7 (5–9)	7 (5–7)	5 (4–6)	-
Hemodynamic parameters				-
PAPm (mm Hg) <sup>‡</sup>	23.8 ± 2.1	51.2 ± 5.5	61.8 ± 13.9	-
CI (L × min <sup>-1</sup> × m <sup>-2</sup> ) <sup>‡</sup>	3.2 ± 0.5	1.9 ± 0.5	2.3 ± 0.3	-
PVR (Wood unit) <sup>‡</sup>	1.8 ± 0.5	11.8 ± 2.2	14.6 ± 3.2	-
Biochemical parameter				-
NT-proBNP (ng × L <sup>-1</sup> ) <sup>‡</sup>	83 ± 60	2,085 ± 924	1,516 ± 1,619	-
Radiologic parameter				-
Lung tissue attenuation (HU) <sup>‡</sup>	- 882.2 ± 24.3	- 828.0 ± 45.1	- 771.3 ± 100.3	- 780.0 ± 44.3

Data are shown as mean ± SD or median with range

\*Measured at rest without or with lowest as tolerable nasal oxygen flow rate

<sup>†</sup> Via nasal cannula

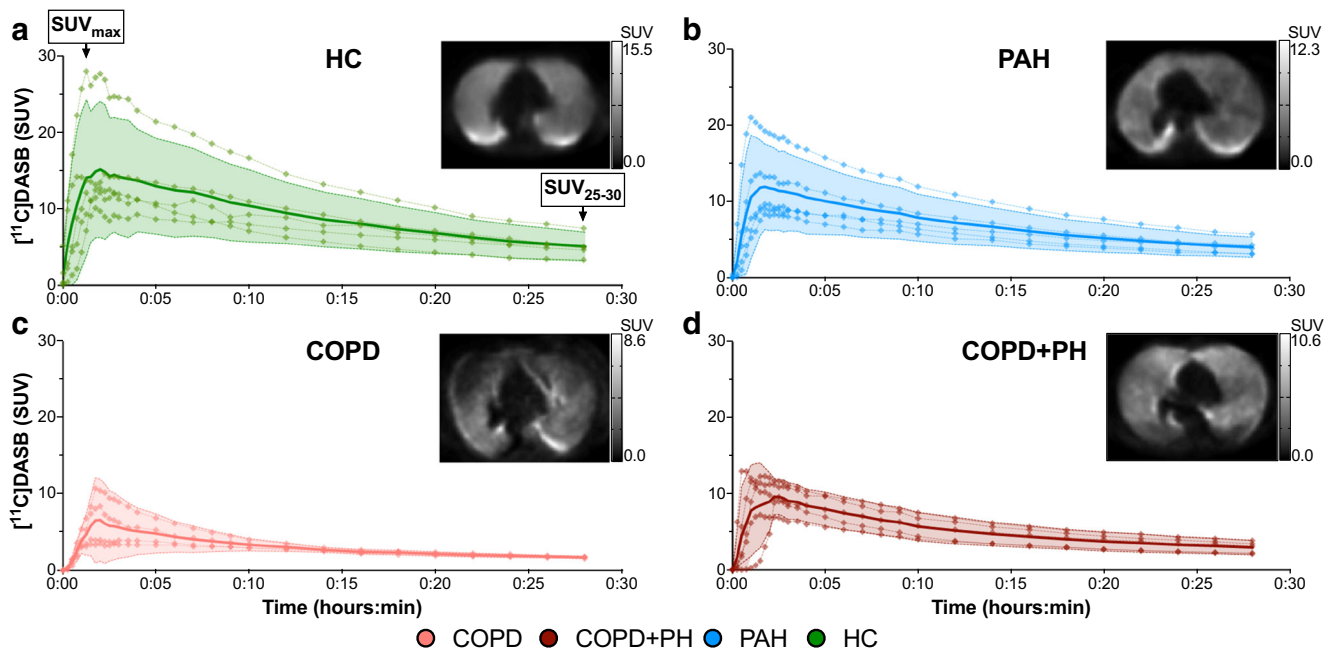
<sup>‡</sup> Significant differences (*P* < 0.05) using ANOVA for > 2 groups or Mann-Whitney *U* test for 2 groups

6MWD 6-min walking distance, A-aO<sub>2</sub> alveolar–arterial gradient, BODE risk of death estimate for COPD patients, CI cardiac index, COPD chronic obstructive pulmonary disease, DLCO diffusion capacity for carbon monoxide after a single breath, FEV<sub>1</sub> forced expiratory volume in 1 s, FiO<sub>2</sub> inspiratory oxygen fraction, FVC forced vital capacity, HC healthy control, HU Hounsfield unit, *n* number of participants, NT-proBNP N-terminal pro-brain natriuretic peptide, PaCO<sub>2</sub> arterial carbon dioxide tension, PAH pulmonary arterial hypertension, PaO<sub>2</sub> arterial oxygen tension, PAPm mean pulmonary arterial pressure, PH pulmonary hypertension, PVR pulmonary vascular resistance, *R*<sub>tot</sub> total airway resistance, RV residual volume, SaO<sub>2</sub> arterial oxygen saturation, TLC total lung capacity, WHO-FC functional classification of pulmonary hypertension according to the World Health Organization

pulmonary SERT availability was found significantly reduced in long-term smokers compared to non-smokers.

The serotonin hypothesis of PH has been proposed for many decades due to serotonin's vasoconstrictive and proliferative properties [4]. The lung has been shown to play a

major role in the physiological removal of circulating serotonin through the presence of pulmonary SERT [11–13]. The SERT-selective tracer [<sup>11</sup>C]DASB and PET imaging studies successfully demonstrated pulmonary SERT availability in humans [18]. There are other SERT-selective



**Fig. 2** Time activity curves of pulmonary  $[^{11}\text{C}]\text{DASB}$  uptake measured within the middle lobe. The time course of  $[^{11}\text{C}]\text{DASB}$  uptake is made of the individual measuring points of each participant and is overlaid with mean values  $\pm 95\%$  confidence interval. The labelling of the axes applies to all panels within the figure, respectively. Each panel contains a representative transversal PET image including the middle lobe at 25-

to 30-min post injection, and a black-white scale showing the range of SUV values. *CI* confidence interval, *COPD* chronic obstructive pulmonary disease, *HC* healthy control, *PAH* pulmonary arterial hypertension, *PH* pulmonary hypertension, *SUV* standardized uptake value, *VOI* volume of interest

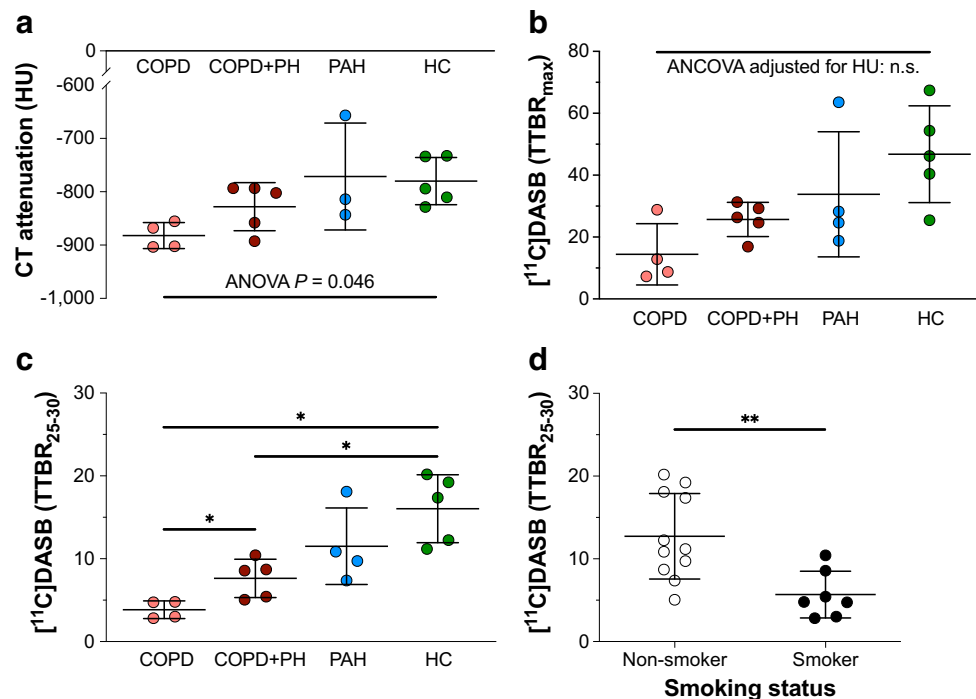
radioligands available for humans as well [23]. Here, we chose to apply  $[^{11}\text{C}]\text{DASB}$  because it binds with high affinity and selectivity to the SERT [24, 25], proving to be suitable for the pulmonary SERT quantification [18, 26]. We provide evidence that pulmonary SERT can be visualized and quantified in patients with COPD and/or PH using  $[^{11}\text{C}]\text{DASB}$  and PET/CT imaging.

Smoke-induced chronic inflammation response leads to parenchymal lung tissue destruction and emphysema, representing a hallmark of advanced COPD stage [3]. As a consequence thereof, pulmonary vascular bed becomes rarefied which affects the pulmonary perfusion. The middle lobe was selected for  $[^{11}\text{C}]\text{DASB}$  uptake analyses since smoke-induced centrilobular emphysema has a typical apical distribution and affects less commonly the middle lobe. Nevertheless, we found significant differences in the variances of lung tissue attenuation values (Fig. 3(a)), which is suggestive of emphysematous and, thus, vascular changes of the lung parenchyma. Likewise, the peak values of  $[^{11}\text{C}]\text{DASB}$  uptake corrected for blood radioactivity ( $\text{TPPR}_{\text{max}}$ ,  $\text{TTBR}_{\text{max}}$ ), likely representing pulmonary perfusion and radiotracer delivery, significantly varied between the cohorts (Fig. 3(b)). These differences vanished not until  $\text{TTBR}_{\text{max}}$  values were corrected for lung tissue attenuation suggesting that pulmonary  $[^{11}\text{C}]\text{DASB}$  uptake was not different between the 4 cohorts.

The physical and biological half-lives of  $[^{11}\text{C}]\text{DASB}$  in humans approximately amount to 20 and 51 min,

respectively. The mean residence time of the tracer is measured 7.6 min, while less than 10% of injected dose are found in the lungs of HC after 30 min of administration [18]. We carried out  $[^{11}\text{C}]\text{DASB}$  uptake analyses between 25 and 30 min p.i. and corrected SUV for both blood and plasma radioactivity ( $\text{TPPR}_{25-30}$ ,  $\text{TTBR}_{25-30}$ ) as well as for lung tissue attenuation. Thereby, we consider the measurement of pulmonary  $[^{11}\text{C}]\text{DASB}$  uptake 25 to 30 min p.i. to be radiotracer-specific for SERT quantification.

It is not possible to apply a compartment model for DASB in the lung. DASB does produce many metabolites [24, 26, 27], which we did not measure in this study, but are yet necessary to obtain an arterial input function for subsequent kinetic modelling. Also, these metabolites are transported into the lung. Thus, the blood and tissue radioactivity measured represents the sum of the parent compound (DASB) and its resulting metabolites, which were not taken into account in this study. The  $\text{TTBR}$  and  $\text{TPPR}$  of  $[^{11}\text{C}]\text{DASB}$ , however, represent a rough approximation of the distribution volume and therefore receptor density in the lung. Furthermore, the lung has a dual blood supply provided by the pulmonary and bronchial circulation, making compartment model assumptions and kinetic modelling of the  $[^{11}\text{C}]\text{DASB}$  and its metabolites very complex. Altogether, these considerations led us to favor a model-free analysis of  $[^{11}\text{C}]\text{DASB}$  uptake in the lung by selecting time frames of interest, which were assumed to



**Fig. 3** Group comparisons of lung tissue attenuation and pulmonary  $[^{11}\text{C}]\text{DASB}$  uptake between the cohorts. (a) Distribution of lung tissue attenuation values among the cohorts measured in HU derived from CT imaging. Overall differences were calculated by means of ANOVA. (b) Group comparison of maximum pulmonary  $[^{11}\text{C}]\text{DASB}$  uptake between the cohorts: After correction for blood activity ( $\text{TTBR}_{\text{max}}$ ) and for attenuation values by means of ANCOVA, the maximum pulmonary  $[^{11}\text{C}]\text{DASB}$  uptake, i.e., pulmonary blood flow, did not differ between the cohorts. (c) Group comparisons of  $[^{11}\text{C}]\text{DASB}$  uptake between the cohorts included the correction for blood activity ( $\text{TTBR}_{25-30}$ ) and for

lung tissue attenuation by means of ANCOVA. (d) Group differences in  $\text{TTBR}_{25-30}$  of  $[^{11}\text{C}]\text{DASB}$  in relation to smoking status. Statistical significance was accepted at a level of a two-sided  $P < 0.05$ . Statistically non-significant comparisons are not labelled,  $*P < 0.05$ ,  $**P < 0.01$ . Group differences are shown by mean  $\pm$  standard deviation. ANCOVA analysis of covariance, ANOVA analysis of variance, COPD chronic obstructive pulmonary disease, HC healthy control, n.s. not significant, PAH pulmonary arterial hypertension, PH pulmonary hypertension, SD standard deviation, TTBR tissue-to-blood concentration ratio

coincide with the tissue to blood equilibrium of the radiotracer in all subjects [17].

To correct the pulmonary  $[^{11}\text{C}]\text{DASB}$  uptake for blood activity, we included the activity measured in both plasma and blood samples. Statistical analysis of the resulting ratios (TPPR, TTBR) showed nearly consistent results.

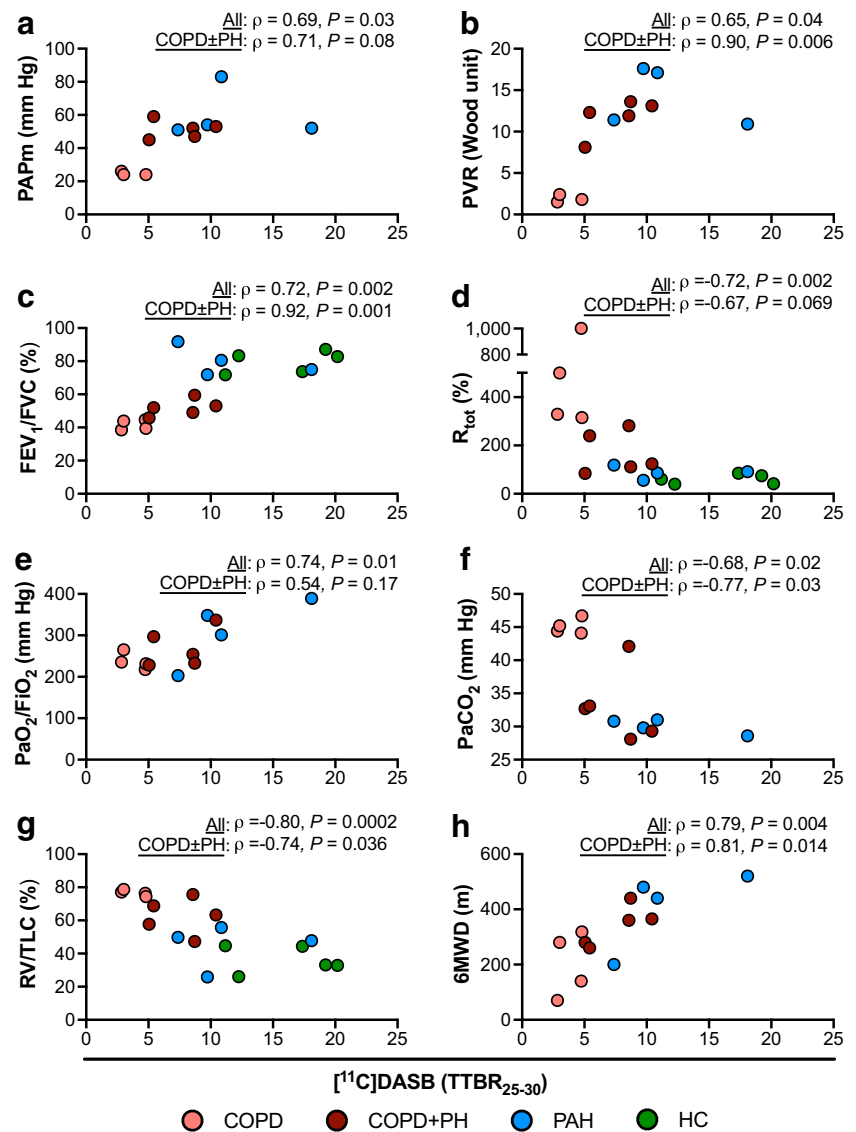
There was no increase in pulmonary SERT availability measured using  $[^{11}\text{C}]\text{DASB}$  in PAH patients compared to HC. This finding contrasts with results from previous investigations that have found an overexpression of SERT in PASM C from PAH patients [5, 6, 28]. In the present study, the PAH cohort consisted of patients with relevant clinical (median WHO-FC III) and hemodynamic impairment (PAPm  $61.8 \pm 13.9$  mm Hg) due to PH. Compared to the PAH patient's PAPm of the 3 aforementioned studies ( $62.0 \pm 13.0$ ,  $62.6 \pm 3.9$ , and  $63 \pm 11.0$  mm Hg, respectively) performing an ex post ANOVA, there was no overall difference in the variances between these 4 studies ( $F = 0.023$ , overall  $P = 0.99$ ). Thus, the difference in SERT expression or  $[^{11}\text{C}]\text{DASB}$  availability cannot be attributed to the severity of PH. Likewise, the PAH and COPD+PH cohort in the present study did not differ in the clinical and hemodynamic

impairment as well as in the  $\text{TTBR}_{25-30}$  values that were corrected for lung attenuation (Table 1).

Notably, COPD and COPD+PH patients who were altogether hypoxemic ( $\text{SaO}_2$ ,  $\text{PaO}_2/\text{FiO}_2$ ) and did neither differ in the degree of airway limitation ( $\text{FEV}_1$ ) nor in the degree of emphysema ( $\text{RV}/\text{TLC}$ ) (Table 1, Supplementary Fig. 1), were measured significantly lower  $\text{TTBR}_{25-30}$  values corrected for lung attenuation than those in the HC. Consistent with this observation is the fact that several rodent models indicate that chronic exposure to hypoxia not only leads to reduced serotonin uptake into the lung [29, 30], but also to downregulation of SERT mRNA or protein expression in PASM C [31–33]. Thus, it may be plausible that hypoxemic COPD patients showed a reduced pulmonary SERT availability in PET/CT using  $[^{11}\text{C}]\text{DASB}$ . However, these reports are challenged by other investigations showing that hypoxia increases SERT expression in PASM C, both in 2 rodent models [34, 35] and in 1 human cohort of COPD+PH patients [14]. MacLean and colleagues argued [32] that the variation in the results of these studies might be due to age, sex, or strain differences of the rodents combined with differences in hypoxic exposure and/or resident atmospheric pressures. For instance, only female



**Fig. 4** Scatter plots of [ $^{11}\text{C}$ ]DASB uptake and hemodynamic (a, b) and clinical parameters (c–h). Partial Spearman rank correlation analysis was performed to correct for lung tissue attenuation. Results are depicted as Spearman rank correlation coefficient rho ( $\rho$ ). Statistical significance was accepted at a two-sided  $P < 0.05$ . The labelling of the abscissa applies to all panels. 6MWD 6-min walking distance, COPD chronic obstructive pulmonary disease, FEV<sub>1</sub> forced expiratory volume in 1 s, FVC forced vital capacity, HC healthy control, PaO<sub>2</sub>/FiO<sub>2</sub> oxygenation ratio: arterial oxygen tension/inspiratory oxygen fraction, PAH pulmonary arterial hypertension, PAPm mean pulmonary arterial pressure, PaCO<sub>2</sub> arterial oxygen tension, PH pulmonary hypertension, PVR pulmonary vascular resistance, RV residual volume, TLC total lung capacity, TTBR<sub>25–30</sub> tissue-to-blood concentration ratio after 25 to 30 minutes of [ $^{11}\text{C}$ ]DASB administration



but not male mice, both overexpressing pulmonary SERT, develop PAH. Their female sex hormone 17 $\beta$  oestradiol is found to induce PAMSC proliferation in the presence of serotonin via stimulating the 5-HT<sub>1B</sub> receptor [36]. In the present study, there was an uneven gender distribution across the 3 patient cohorts consisting of predominantly male participants in the COPD and COPD+PH cohort (4/5 male, respectively), while exclusively female participants were in the PAH cohort. One cannot rule out the possibility that the gender distribution influenced the SERT availability, which, nevertheless, was balanced among COPD patients with or without PH.

Taken together, since SERT availability in the present study was reduced in hypoxemic COPD patients (Fig. 3(c), Table 1), a hypoxia-induced downregulation of SERT could explain our findings.

Even though COPD patients with or without PH expressed reduced pulmonary SERT availability compared to HC, there

was still a significant increase in SERT availability when COPD patients developed PH (Fig. 3(c)). This suggests, that the upregulation of SERT may play a potential role in the development of PH in COPD patients.

In addition to the quantity of functional SERT protein in the cytoplasmic membrane of PAMSC, the transporter activity seems to influence the uptake of serotonin [6]. Furthermore, risk of developing PH and its severity in the course of COPD appears to be associated with certain SERT gene polymorphisms, especially with the LL genotype of its promoter region [37, 38]. Accumulating evidence suggests that 5-HT receptors regulate the SERT activity in the PAMSC membrane leading to lower serotonin uptake and in turn higher extracellular serotonin levels available for 5-HT receptor activation [30, 33].

In contrast to the constricting action of serotonin on PAMSC, which is mainly mediated by 5-HT<sub>1B</sub> receptors,

**Table 2** Partial correlation of TTBR<sub>25–30</sub> with pulmonary function and biochemical and hemodynamic parameters with correction for lung tissue attenuation

		Airflow limitation			Emphysema	Gas exchange				Exercise capacity	Right heart strain	Hemodynamic		
		FEV <sub>1</sub> /FVC	FEV <sub>1</sub>	R <sub>tot</sub>		RV/TLC	DLCO	PaO <sub>2</sub> /F <sub>i</sub> O <sub>2</sub>	SaO <sub>2</sub>			PaCO <sub>2</sub>	6MWD	NT-proBNP
All*	$\rho$	0.72	0.73	− 0.71	− 0.80	0.77	0.74	0.69	− 0.68	0.79	0.07	0.69	0.65	− 0.36
	P	0.002	0.001	0.002	0.0002	0.01	0.01	0.02	0.02	0.004	0.85	0.03	0.04	0.31
	n	17	17	17	17	11	12	12	12	12	12	11	11	11
COPD ± PH	$\rho$	0.92	0.73	− 0.67	− 0.74	0.42	0.54	0.45	− 0.77	0.81	0.83	0.71	0.90	− 0.92
	P	0.001	0.04	0.069	0.036	0.35	0.17	0.26	0.03	0.014	0.01	0.075	0.006	0.003
	n	9	9	9	9	8	9	9	9	9	9	8	8	8

\*Taking into account the data available from the 4 cohorts, e.g., there were no measurements of gas exchange and hemodynamic in HC. Partial Spearman rank correlation (*ρ*) of TTBR<sub>25–30</sub> with pulmonary function and biochemical and hemodynamic parameters was used to correct for lung tissue attenuation of the middle lobe measured by computed tomography between the 4 cohorts (i.e., all) and between the COPD cohort with or without PH (i.e., COPD ± PH). Statistically significant partial correlations were accepted at a level of a two-sided *P* < 0.05 and are reported in italic

6MWD 6-min walking distance, BGA blood gas analysis, CI cardiac index, DLCO diffusion capacity for carbon monoxide after a single breath, FEV<sub>1</sub> forced expiratory volume in 1 s, FiO<sub>2</sub> inspiratory oxygen fraction, FVC forced vital capacity, *n* number of participants, NT-proBNP N-terminal pro-brain natriuretic peptide, PaCO<sub>2</sub> carbon dioxide tension in arterial blood, PaO<sub>2</sub> oxygen tension in arterial blood, PAPm mean pulmonary arterial pressure, PFT pulmonary function test, PH pulmonary hypertension, PVR pulmonary vascular resistance, R<sub>tot</sub> total airway resistance, RV residual volume, TLC total lung capacity, TTBR<sub>25–30</sub> tissue-to-blood concentration ratio of pulmonary [<sup>11</sup>C]DASB uptake

the proliferative effects of serotonin require its internalization by the SERT [4, 34]. Serotonin that is taken up by the SERT activates not only mitogen-activated protein kinases representing proliferative stimuli, but also activates nicotinamide adenine dinucleotide phosphate (NADPH) oxidases leading to increased generation of reactive oxygen species (ROS), such as hydrogen peroxide. The subunit of NADPH oxidases, p22phox, is believed to play a role as a hypoxia sensor for HPV. In hypoxemic COPD patients, p22phox positively correlates with PAPm and oxygenation ratio as well as negatively with gas exchange capacity, suggesting that NADPH oxidases may facilitate HPV [39]. In COPD patients with or without PH, we found that the quantity of SERT availability, which can be considered as a surrogate for the quantity of internalized serotonin and thus potential inducer of NADPH oxidases (p22phox), did not correlate with oxygenation ratio or gas exchange capacity (DLCO). But it positively correlated with severity of PH (PVR), right heart strain (NT-proBNP), and airflow limitation (Fig. 4, Table 2), indicating a potential pathophysiologic link between pulmonary SERT availability, NADPH oxidases, ROS production, and thus HPV in COPD patients with or without PH.

Limitations of this study comprise the low number of participants, since for thorough analysis of effect size, a larger population will certainly be needed. Even though the gender distribution between the COPD cohorts including patients with or without PH was balanced (4 men, 1 woman, respectively), the control groups showed a different distribution (exclusively women in the PAH cohort, 2 men and 3 women in

the HC). From these distribution differences, we cannot exclude the influence of gender-specific differences when comparing COPD cohort with PAH or healthy cohorts. Furthermore, we did not apply a compartment model for estimating binding potential using kinetic modelling of the [<sup>11</sup>C]DASB. We applied a semiquantitative approach to measure pulmonary [<sup>11</sup>C]DASB uptake by using the tissue-to-blood-concentration ratio after 25 to 30 min of tracer injection. We were aware that when selecting a time frame of interest, this should coincide with the transient equilibrium of the tracer in all subjects [17]. The HC did not undergo RHC, blood gas analysis, and exercise capacity.

In conclusion, pulmonary SERT availability was demonstrated using [<sup>11</sup>C]DASB and PET/CT in COPD patients with or without PH, in PAH patients as well as in HC. COPD patients, regardless of the presence of PH, showed reduced [<sup>11</sup>C]DASB uptake, while COPD patients who developed PH were found increased [<sup>11</sup>C]DASB uptake. These findings suggest that pulmonary SERT availability may reflect an important aspect in the pathogenesis of PH in patients with COPD, and its measurement may form the basis for further developments in the diagnosis and management of PH in COPD patients.

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## Compliance with ethical standards

**Conflicts of interest/competing interests** The authors declare that they have no conflict of interest.

**Ethics approval** This study received ethical approval from the local ethics committee at the Medical Faculty, University of Leipzig (IRB00001750, AZ: 207/13) and was approved by the regulatory authorities in Germany (Bundesamt für Strahlenschutz, BfS, Federal Office for Radiation Protection, ZS-22463/2-2014-017). All procedures performed in this study involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

**Informed consent** All participants gave their written informed consent prior to study enrolment and for publication.

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## References

- Galie N, Humbert M, Vachiery JL, Gibbs S, Lang I, Torbicki A, et al. 2015 ESC/ERS Guidelines for the diagnosis and treatment of pulmonary hypertension: the joint task force for the diagnosis and treatment of pulmonary hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS). *Eur Respir J*. 2015;46(4):903–75. <https://doi.org/10.1183/13993003.01032-2015>.
- Seeger W, Adir Y, Barbera JA, Champion H, Coghlan JG, Cottin V, et al. Pulmonary hypertension in chronic lung diseases. *J Am Coll Cardiol*. 2013;62(25 Suppl):D109–16. <https://doi.org/10.1016/j.jacc.2013.10.036>.
- Global initiative for chronic obstructive lung disease. Global strategy for diagnosis, management and prevention of COPD (2020 Report). [www.goldcopd.org](http://www.goldcopd.org). 2019. Accessed 16 Nov 2019.
- MacLean MMR. The serotonin hypothesis in pulmonary hypertension revisited: targets for novel therapies (2017 Grover Conference Series). *Pulm Circ*. 2018;8(2):2045894018759125. <https://doi.org/10.1177/2045894018759125>.
- Eddahibi S, Guignabert C, Barlier-Mur AM, Dewachter L, Fadel E, Dartevielle P, et al. Cross talk between endothelial and smooth muscle cells in pulmonary hypertension: critical role for serotonin-induced smooth muscle hyperplasia. *Circulation*. 2006;113(15):1857–64. <https://doi.org/10.1161/CIRCULATIONAHA.105.591321>.
- Eddahibi S, Humbert M, Fadel E, Raffestin B, Darmon M, Capron F, et al. Serotonin transporter overexpression is responsible for pulmonary artery smooth muscle hyperplasia in primary pulmonary hypertension. *J Clin Invest*. 2001;108(8):1141–50. <https://doi.org/10.1172/JCI12805>.
- Guignabert C, Izikki M, Tu LI, Li Z, Zadigue P, Barlier-Mur AM, et al. Transgenic mice overexpressing the 5-hydroxytryptamine transporter gene in smooth muscle develop pulmonary hypertension. *Circ Res*. 2006;98(10):1323–30. <https://doi.org/10.1161/01.RES.0000222546.45372.a0>.
- Morecroft I, Pang L, Baranowska M, Nilsen M, Loughlin L, Dempsey Y, et al. In vivo effects of a combined 5-HT<sub>1B</sub> receptor/SERT antagonist in experimental pulmonary hypertension. *Cardiovasc Res*. 2010;85(3):593–603. <https://doi.org/10.1093/cvr/cvp306>.
- Sadoughi A, Roberts KE, Preston IR, Lai GP, McCollister DH, Farber HW, et al. Use of selective serotonin reuptake inhibitors and outcomes in pulmonary arterial hypertension. *Chest*. 2013;144(2):531–41. <https://doi.org/10.1378/chest.12-2081>.
- Graham D, Langer SZ. Advances in sodium-ion coupled biogenic amine transporters. *Life Sci*. 1992;51(9):631–45. [https://doi.org/10.1016/0024-3205\(92\)90236-i](https://doi.org/10.1016/0024-3205(92)90236-i).
- Strum JM, Junod AF. Radioautographic demonstration of 5-hydroxytryptamine-3 H uptake by pulmonary endothelial cells. *J Cell Biol*. 1972;54(3):456–67. <https://doi.org/10.1083/jcb.54.3.456>.
- Junod AF. Uptake, metabolism and efflux of 14 C-5-hydroxytryptamine in isolated perfused rat lungs. *J Pharmacol Exp Ther*. 1972;183(2):341–55.
- Paczkowski NJ, Vuocolo HE, Bryan-Lluka LJ. Conclusive evidence for distinct transporters for 5-hydroxytryptamine and noradrenaline in pulmonary endothelial cells of the rat. *Naunyn Schmiedeberg's Arch Pharmacol*. 1996;353(4):423–30. <https://doi.org/10.1007/bf00261439>.
- Eddahibi S, Chaouat A, Morrell N, Fadel E, Fuhrman C, Bugnet AS, et al. Polymorphism of the serotonin transporter gene and pulmonary hypertension in chronic obstructive pulmonary disease. *Circulation*. 2003;108(15):1839–44. <https://doi.org/10.1161/01.CIR.0000091409.53101.E8>.
- Eddahibi S, Hanoun N, Lanfumey L, Lesch KP, Raffestin B, Hamon M, et al. Attenuated hypoxic pulmonary hypertension in mice lacking the 5-hydroxytryptamine transporter gene. *J Clin Invest*. 2000;105(11):1555–62. <https://doi.org/10.1172/JCI8678>.
- Suhara T, Sudo Y, Yoshida K, Okubo Y, Fukuda H, Obata T, et al. Lung as reservoir for antidepressants in pharmacokinetic drug interactions. *Lancet*. 1998;351(9099):332–5. [https://doi.org/10.1016/S0140-6736\(97\)07336-4](https://doi.org/10.1016/S0140-6736(97)07336-4).
- Norgaard M, Ganz M, Svarer C, Feng L, Ichise M, Lanzenberger R, et al. Cerebral serotonin transporter measurements with [<sup>11</sup>C]DASB: A review on acquisition and preprocessing across 21 PET centres. *J Cereb Blood Flow Metab*. 2019;39(2):210–22. <https://doi.org/10.1177/0271678X18770107>.
- Lu JQ, Ichise M, Liow JS, Ghose S, Vines D, Innis RB. Biodistribution and radiation dosimetry of the serotonin transporter ligand <sup>11</sup>C-DASB determined from human whole-body PET. *J Nucl Med*. 2004;45(9):1555–9.

19. Booij J, de Jong J, de Bruin K, Knol R, de Win MM, van Eck-Smit BL. Quantification of striatal dopamine transporters with  $^{123}\text{I}$ -FP-CIT SPECT is influenced by the selective serotonin reuptake inhibitor paroxetine: a double-blind, placebo-controlled, crossover study in healthy control subjects. *J Nucl Med*. 2007;48(3):359–66.
20. Vogelmeier CF, Criner GJ, Martinez FJ, Anzueto A, Barnes PJ, Bourbeau J, et al. Global strategy for the diagnosis, management, and prevention of chronic obstructive lung disease 2017 report: GOLD executive summary. *Eur Respir J*. 2017;49(3). <https://doi.org/10.1183/13993003.00214-2017>.
21. Celli BR, Cote CG, Marin JM, Casanova C, Montes de Oca M, Mendez RA, et al. The body-mass index, airflow obstruction, dyspnea, and exercise capacity index in chronic obstructive pulmonary disease. *N Engl J Med*. 2004;350(10):1005–12. <https://doi.org/10.1056/NEJMoa021322>.
22. Barst RJ, McGoon M, Torbicki A, Sitbon O, Krowka MJ, Olschewski H, et al. Diagnosis and differential assessment of pulmonary arterial hypertension. *J Am Coll Cardiol*. 2004;43(12 Suppl S):40S–7S. <https://doi.org/10.1016/j.jacc.2004.02.032>.
23. Huang Y, Zheng MQ, Gerdes JM. Development of effective PET and SPECT imaging agents for the serotonin transporter: has a twenty-year journey reached its destination? *Curr Top Med Chem*. 2010;10(15):1499–526. <https://doi.org/10.2174/156802610793176792>.
24. Wilson AA, Ginovart N, Schmidt M, Meyer JH, Threlkeld PG, Houle S. Novel radiotracers for imaging the serotonin transporter by positron emission tomography: synthesis, radiosynthesis, and in vitro and ex vivo evaluation of  $^{11}\text{C}$ -labeled 2-(phenylthio)araalkylamines. *J Med Chem*. 2000;43(16):3103–10. <https://doi.org/10.1021/jm000079i>.
25. Frankle WG, Huang Y, Hwang DR, Talbot PS, Slifstein M, Van Heertum R, et al. Comparative evaluation of serotonin transporter radioligands  $^{11}\text{C}$ -DASB and  $^{11}\text{C}$ -McN 5652 in healthy humans. *J Nucl Med*. 2004;45(4):682–94.
26. Parsey RV, Ojha A, Ogden RT, Erlandsson K, Kumar D, Landgrebe M, et al. Metabolite considerations in the in vivo quantification of serotonin transporters using  $^{11}\text{C}$ -DASB and PET in humans. *J Nucl Med*. 2006;47(11):1796–802.
27. Ginovart N, Wilson AA, Meyer JH, Hussey D, Houle S. Positron emission tomography quantification of  $[(^{11}\text{C})\text{-DASB}]$  binding to the human serotonin transporter: modeling strategies. *J Cereb Blood Flow Metab*. 2001;21(11):1342–53. <https://doi.org/10.1097/00004647-200111000-00010>.
28. Marcos E, Fadel E, Sanchez O, Humbert M, Darteville P, Simonneau G, et al. Serotonin-induced smooth muscle hyperplasia in various forms of human pulmonary hypertension. *Circ Res*. 2004;94(9):1263–70. <https://doi.org/10.1161/01.RES.0000126847.27660.69>.
29. Jeffery TK, Bryan-Lluka LJ, Wanstall JC. Specific uptake of 5-hydroxytryptamine is reduced in lungs from hypoxic pulmonary hypertensive rats. *Eur J Pharmacol*. 2000;396(2-3):137–40. [https://doi.org/10.1016/s0014-2999\(00\)00252-1](https://doi.org/10.1016/s0014-2999(00)00252-1).
30. Launay JM, Herve P, Peoc'h K, Tournois C, Callebort J, Nebigil CG, et al. Function of the serotonin 5-hydroxytryptamine 2B receptor in pulmonary hypertension. *Nat Med*. 2002;8(10):1129–35. <https://doi.org/10.1038/nm764>.
31. Hoshikawa Y, Nana-Sinkam P, Moore MD, Sotto-Santiago S, Phang T, Keith RL, et al. Hypoxia induces different genes in the lungs of rats compared with mice. *Physiol Genomics*. 2003;12(3):209–19. <https://doi.org/10.1152/physiolgenomics.00081.2001>.
32. MacLean MR, Deuchar GA, Hicks MN, Morecroft I, Shen S, Sheward J, et al. Overexpression of the 5-hydroxytryptamine transporter gene: effect on pulmonary hemodynamics and hypoxia-induced pulmonary hypertension. *Circulation*. 2004;109(17):2150–5. <https://doi.org/10.1161/01.CIR.0000127375.56172.92>.
33. Morecroft I, Loughlin L, Nilsen M, Colston J, Dempsey Y, Sheward J, et al. Functional interactions between 5-hydroxytryptamine receptors and the serotonin transporter in pulmonary arteries. *J Pharmacol Exp Ther*. 2005;313(2):539–48. <https://doi.org/10.1124/jpet.104.081182>.
34. Eddahibi S, Fabre V, Boni C, Martres MP, Raffestin B, Hamon M, et al. Induction of serotonin transporter by hypoxia in pulmonary vascular smooth muscle cells. Relationship with the mitogenic action of serotonin. *Circ Res*. 1999;84(3):329–36. <https://doi.org/10.1161/01.res.84.3.329>.
35. Ciuculan L, Hussey MJ, Burton V, Good R, Duggan N, Beach S, et al. Imatinib attenuates hypoxia-induced pulmonary arterial hypertension pathology via reduction in 5-hydroxytryptamine through inhibition of tryptophan hydroxylase 1 expression. *Am J Respir Crit Care Med*. 2013;187(1):78–89. <https://doi.org/10.1164/rccm.201206-1028OC>.
36. White K, Dempsey Y, Nilsen M, Wright AF, Loughlin L, MacLean MR. The serotonin transporter, gender, and 17beta oestradiol in the development of pulmonary arterial hypertension. *Cardiovasc Res*. 2011;90(2):373–82. <https://doi.org/10.1093/cvr/cvq408>.
37. Zhang H, Xu M, Xia J, Qin RY. Association between serotonin transporter (SERT) gene polymorphism and idiopathic pulmonary arterial hypertension: a meta-analysis and review of the literature. *Metabolism*. 2013;62(12):1867–75. <https://doi.org/10.1016/j.metabol.2013.08.012>.
38. Jiao YR, Wang W, Lei PC, Jia HP, Dong J, Gou YQ, et al. 5-HTT, BMPR2, EDN1, ENG, KCNA5 gene polymorphisms and susceptibility to pulmonary arterial hypertension: A meta-analysis. *Gene*. 2019;680:34–42. <https://doi.org/10.1016/j.gene.2018.09.020>.
39. Nagaraj C, Tabeling C, Nagy BM, Jain PP, Marsh LM, Papp R, et al. Hypoxic vascular response and ventilation/perfusion matching in end-stage COPD may depend on p22phox. *Eur Respir J*. 2017;50(1). <https://doi.org/10.1183/13993003.01651-2016>.

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